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Original Article

The Hemoglobin Levels and Transfusion Intervals of Beta-Thalassemia Patients with Positive and Negative Allo-Autoantibodies

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ABSTRACT

Beta-thalassemia is a common cause of anemia. Blood transfusion is the primary therapy for beta-thalassemia patients in Indonesia who are at risk of developing allo-autoantibodies. These antibodies can lead to the lysis of red blood cells, resulting in a rapid decrease in hemoglobin levels and shorter transfusion intervals. This research aims to compare hemoglobin levels and transfusion intervals in beta-thalassemia patients with positive and negative allo-autoantibodies. This research is a retrospective cohort study utilizing medical record data from beta-thalassemia patients who underwent blood transfusions at Dr. Soetomo Hospital. The patients were divided into two groups: those with positive allo-autoantibodies and those with negative allo-autoantibodies. Data on hemoglobin levels and transfusion dates were collected five times consecutively between July and December, 2021. The hemoglobin levels of the two groups were compared using the Mann-Whitney test, while the transfusion intervals were analysed using the T-test. Data were obtained from 52 beta-thalassemia patients who received transfusions, with 25 (48%) testing positive for alloautoantibodies and 27 (52%) testing negative. It was observed that the hemoglobin levels of the two groups were not significantly different (p = 0.769). Similarly, the transfusion intervals of the two groups were not significantly different (p = 0.899). There were no significant differences in hemoglobin levels and transfusion intervals between patients with positive and negative allo-autoantibodies.

Background

- Beta-thalassemia as a
- common cause of anemia.
- Blood transfusion as the primary therapy
- Formation of alloautoantibodies leading to
- red blood cell lysis.



Aim Comparing hemoglobin levels and transfusion intervals in betathalassemia patients with positive and negative alloautoantibodies

GRAPHICALABSTRACT

Result

Patients with positive and negative allo-autoantibodies did not differ in their hemoglobin levels or intervals between transfusions

Introduction

Beta-thalassemia stands out as a prevalent cause of anemia, originating from a diminished or absent synthesis of the beta-globin chains of hemoglobin [1]. It represents the most common inherited single-gene abnormality [2]. Indonesia, being part of the global thalassemia belt, exhibits a high frequency of the thalassemia gene, ranging from 3-10% [3]. Reports from the Eijkman National Molecular Institute indicate a carrier gene frequency of beta-thalassemia in Indonesia within this range [4]. As of 2019, the estimated number of beta-thalassemia patients in Indonesia reached approximately 10,000 [5]. The current approach to managing beta-thalassemia is predominantly symptomatic, involving lifelong blood transfusions [6]. For beta-thalassemia patients, the indication for transfusion is a hemoglobin level below 7 g/dL in two consecutive examinations, with an interval exceeding two weeks and without signs of infection. Alternatively, a hemoglobin level above 7 g/dL may prompt transfusion if there is evidence of failure to thrive or bone deformities resulting from thalassemia [7].

The targeted pre-transfusion hemoglobin level is set at 9-10 g/dL, while post-transfusion levels aim for 13-14 g/dL. The typical transfusion interval for beta-thalassemia patients ranges from 2 to 4 weeks [8]. Repeated blood transfusions in transfusion-dependent betathalassemia patients elevate the risk of infectious disease transmission, volume overload, hemolytic transfusion reactions, iron overload, allo- and autoimmunization, and Delayed Haemolytic Transfusion Reactions (DHTRs) [6, 9, 10].

Allo-autoantibodies contribute to the lysis of red blood cells, shortening their lifespan, diminishing pre-transfusion hemoglobin levels, and requiring more frequent transfusion intervals [7, 11]. Kurniawan *et al.*'s research revealed that transfusion-dependent beta-thalassemia patients with allo-autoantibodies failed to maintain hemoglobin levels after transfusion compared to those without allo-autoantibodies. This finding aligns with Essa *et al.*'s research, indicating that transfusion-dependent beta-thalassemia patients with allo-autoantibodies require significantly more frequent transfusions per year [12, 13]. This study aims to compare hemoglobin levels and transfusion intervals in transfusiondependent beta-thalassemia patients, specifically between groups with positive and negative allo-

autoantibodies, at Dr. Soetomo Hospital.

Martials and Methods

This retrospective cohort study was conducted in August-September 2022, with research data obtained subsequent to receiving approval from the Health Research Ethics Committee of Dr. Soetomo Hospital (approval letter number: 1006/LOE/301.4.2/VIII/2022). The study involved the collection of medical record data from beta-thalassemia patients who underwent transfusions at Dr. Soetomo Hospital in Surabaya, East Java, Indonesia. Inclusion criteria comprised patients whose medical records confirmed a diagnosis of beta-thalassemia for minimal six months and who had received a minimum of 5 blood transfusions between July and December 2021. Exclusion criteria encompassed betathalassemia patients who had received fewer than 5 transfusions, and patients with infectious, malignant, or degenerative diseases based on medical records. Hemoglobin data, based on medical record information, was measured using the Sysmex Automated Hematology Analyzer XN-3000 (Sysmex, Kobe, Japan) utilizing the SLS-Hemoglobin method. Alloantibody data were determined through antibody screening in pretransfusion tests using the QWALYS 3 (Diagast, Loos, France) with the solid-phase adherence method. Autoantibody data were assessed based on auto-control results using Diamed gel cards (Bio-Rad, Cressier, Switzerland). The collected data were categorized into two groups: the positive allo-autoantibody group and the negative allo-autoantibody group.

The Kolmogorov-Smirnov test was employed to assess the normality of the research data. Differences in hemoglobin levels between the two groups were analysed using the Mann-Whitney test, while differences in transfusion intervals were analysed using the T-test. The data was processed using SPSS version 25 (IBM Corp., United States), with a significance level set at p < 0.05.

Results and Discussion

Patient characteristics

A total of 52 data sets from transfusiondependent beta-thalassemia patients were analyzed in this study, with 25 patients (48%) in the positive allo-autoantibody group and 27 patients (52%) in the negative allo-autoantibody group. Patient characteristics are summarized in Table 1.

The antibody examination of transfusiondependent beta-thalassemia patients revealed that 10 patients (19.2%) exhibited both alloautoantibodies. In addition, 21 patients (40%) had autoantibodies, while 14 patients (27%) had alloantibodies. Notably, the number of patients exclusively presenting autoantibodies (n = 11) surpassed those with only alloantibodies (n = 4) (Table 2).

Comparison of hemoglobin levels and transfusion intervals

The comparison of hemoglobin levels and transfusion intervals in transfusion-dependent beta-thalassemia patients between the positive and negative allo-autoantibody groups did not yield significant differences (p>0.05), as indicated in Table 3.

Characteristics	Allo-Auto Antibody Positive Allo-Auto Antibody Negativ		
	n (%)	n (%)	
Total	25 (48)	27 (52)	
Gender			
Man	5 (20)	15 (55.6)	
Woman	20 (80)	12 (44.4)	
Age			
≤ 18 years	15 (60)	19 (70.4)	
> 18 years old	10 (40)	8 (29.6)	
Number of transfusions			
≤ 50 times	8 (32)	9 (33.3)	
51-150 times	9 (36)	11 (40.7)	
151-250 times	5 (20)	7 (26)	
251-350 times	3 (12)	0	

Table 1: Patient characteristics

Table 2: The occurrence of allo-autoantibodies

	Alloantibodies		
	Positive n=14 (27%)	Negative	
Autoantibodies			
Positive n=21 (40%)	10	11	
Negative	4	27	

 Table 3: Comparison of hemoglobin levels and transfusion intervals between positive and negative alloautoantibody groups

Variable	Positive allo-autoantibody	Negative allo-autoantibody	<i>P</i> -value
Hemoglobin level	7.9 g/dL	7.92 g/dL	0.769
(Median)			
Transfusion intervals (Mean)	3.31 weeks	3.27 weeks	0.899

In this study, it was observed that the positive allo- and autoantibody group was predominantly composed of women, aligning with findings from research by Essa et al. and Singer et al. These studies suggest that nulliparous women are at a higher risk of experiencing Red Blood Cell (RBC) sensitization [13, 14]. However, the contrary results were obtained from the research by Ameen et al., Dhawan et al., Hendrickson et al., and Saifeldeen et al., which indicated no significant relationship between gender and the incidence of allo- and autoantibodies in transfusion-dependent beta-thalassemia patients [15-18]. The age distribution of patients in our study did not show significant differences between the two groups, aligning with Dhawan et *al.*'s research [16]. The relationship between the number of transfusions and the occurrence of allo- and autoantibodies remains unclear, as suggested by the literature [19]. In our study, the average number of transfusions in both groups was identical, at 100.26 times. This finding is consistent with the research by Dhawan et al., Saifeldeen et al., Ahmed et al., and Obeidi et al. [16, 18, 20, 21]. In contrast, Singer et al. and Vichinsky *et al.* reported that a higher number of transfusions is associated with an increased incidence of allo-autoantibodies [14, 22]. Notably, our study identified three patients with allo- and autoantibodies who had undergone 300 or more transfusions, with ages ranging from 30, 33, to 38 years. This suggests advancements in prevention and management efforts, contributing to an increased life expectancy for transfusiondependent beta-thalassemia patients in Indonesia compared to the situation in 1978 when life expectancy was around 8-10 years [5].

Alloantibodies immune are an response stimulated by repeated PRC (Packed Red Cell) transfusions. Several factors that can cause the formation of alloantibodies are differences in RBC antigens between donor and recipient blood, the recipient's immune status, and the immunomodulatory effect on the recipient's immune system [14, 16, 23]. Previous studies reported the incidence of alloantibodies varying from 4-50% [14, 16, 23, 24]. High incidence is observed when donor and recipient populations heterogeneous, compatibility are testing procedures are lacking, the age at which transfusion begins is > 1 year, nonleucoreduced PRC is administered, and a history of splenectomy exists [7, 14, 16, 18, 23, 24]. Initiating transfusions after the age of 1 year can lead to alloantibody formation due to the loss of protection from the immature immune system and the breakdown of adaptive immune tolerance to allogeneic RBC antigens [16, 19, 24] Administration of nonleucoreduced PRC and a history of splenectomy can also contribute to alloantibody appearance by inducing lymphocytosis associated with blood transfusions, leading to increased serum immunoglobulins, immune complexes, and cells expressing immunoglobulins [14, 16, 25]. This study found a relatively high incidence of alloantibodies in transfusion-dependent betathalassemia patients, specifically at 27%. This aligns closely with the findings of Saifeldeen et al. (23.1%) and Singer *et al.* (22%) [14, 18]. Potential contributors to alloantibody formation in our study include population heterogeneity, a suspected age at which transfusion begins exceeding 1 year, and a suspected history of splenectomy.

In this study, 40% of patients were found to have autoantibodies, which is notably higher than reported in previous research by Dhawan et al. (28.2%), Saifeldeen et al. (9.2%), Singer et al. (25%), Ameen et al. (11%), Pahuja et al. (0.47%), and Noor et al. [14-16, 18, 23, 26]. The factors contributing to the formation of autoantibodies in transfusion-dependent beta-thalassemia patients remain poorly understood. Singer et al. and Dhawan et al. propose a connection between autoantibody formation and a history of splenectomy, suggesting that the loss of an efficient filtration system leads to the exposure of old Red Blood Cells (RBCs) to new antigens, triggering an immune response, including the formation of autoantibodies. However, Noor et al. found no association between the occurrence of autoantibodies and a history of splenectomy [14, 16, 26]. Singer et al. further reported that the incidence of autoantibodies is related to a history of previous alloantibodies and the administration of nonleucoreduced PRC. In contrast, Noor et al. proposed that autoantibody formation can be

influenced by storing PRC at a temperature of 1-6 °C for more than 3 days. This storage condition induces an increase in White Blood Cell (WBC) apoptosis, releasing immunostimulatory antigens and biological mediators (core protein matrix, CTLA-4 epitope), thereby sensitizing the patient's immune system and triggering autoantibody formation [14, 26-29]. The high prevalence of patients with autoantibodies in our study is suspected to be influenced by previous alloantibodies and a history of splenectomy. Remarkably, this study identified 11 patients who exclusively had autoantibodies. The appearance of autoantibodies without preceding alloantibodies might be attributed to immune system dysfunction resulting from genetic factors, T-reg cell responses to red blood cell autoantigen epitopes mediated by IL-10, and severe hemoglobinopathy (HbE/ β thalassemia) [26, 30].

The comparison of hemoglobin levels and transfusion intervals between the two groups in this study did not reveal significant differences. This aligns with the findings of Obeidi et al. in an Iranian population, where no relationship was found between pretransfusion hemoglobin levels, transfusion intervals, and the presence of alloautoantibodies. Similarly, Saifeldeen et al.'s study in Egypt reported no significant difference in transfusion intervals between groups with and without allo-autoantibodies [18, 21]. Contrasting results were observed in Essa's research on an Egyptian population, where the group with alloautoantibodies had shorter transfusion intervals [13]. In addition, Kurniawan *et al.*'s study in Indonesia found that 37.5% of transfusiondependent beta-thalassemia patients failed to maintain post-transfusion hemoglobin levels for 4 weeks, resulting in shorter transfusion intervals (average 23 days). In this group, the alloantibodies prevalence was 78.6%, and autoantibodies were present in 72.7% of cases [12]. These varied outcomes across studies highlight that the presence of alloand autoantibodies may not consistently be a singular factor causing a decrease in pre-transfusion hemoglobin levels and а shortening of transfusion intervals in transfusion-dependent beta-thalassemia patients.

Other factors that may influence these outcomes include patient-related factors (such as history of splenomegaly/splenectomy, comorbidities, type of irregular antibodies possessed, psychology, and financial considerations), blood bank factors (such as storage and availability), and transfusion accuracy factors (such as schedule and quantity). Limitations of this research include its singlelocation focus and limited patient data. Factors other than allo-autoantibodies that can affect hemoglobin levels and transfusion interval still require further research. Researchers suggest the administration of leukoreduced packed red cells (PRC) and antibody screening in pretransfusion testing to mitigate the elevated incidence of alloautoantibodies.

Conclusion

To sum up, the hemoglobin levels and transfusion intervals in transfusion-dependent betathalassemia patients showed no significant differences between the allo- and autoantibody positive and negative groups.

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References

[1]. Needs T., Gonzalez-Mosquera L.F., Lynch D.T., Beta Thalassemia, *StatPearls, Treasure Island: StatPearls*, 2023 [Publisher]

[2]. Rismayanti L., Andarsini M.R., Qibtiyah M., Analysis of DL-α-Tocopherol As Antioxidant On Malondialdehyde Level In Pediatric Patients With β-Thalassemia Major, *Folia Medica Indonesiana*, 2017, **53**:49 [Crossref], [Google Scholar], [Publisher]

[3]. Ministry of Health of the Republic Indonesia., Decree of the Minister of Health of the Republic of Indonesia Number Hk.01.07/Menkes/1/2018 concerning National Guidelines for Medical Services for the Management of Thalassemia 2018 [Publisher]

[4]. Tambunan B.A., Ugrasena I.D.G., Aryati A., Impact of Hemin on Interleukin-21 Levels and Plasma Cells in Transfusion-Dependent Thalassemia with Positive and Negative Allo-Autoantibody, *International Journal of General Medicine*, 2023, **16**:47 [Google Scholar], [Publisher]

[5]. Ministry of Health of the Republic Indonesia., Angka Pembawa Sifat Talasemia Tergolong Tinggi 2019 [<u>Publisher</u>]

[6]. Elhence P., Solanki A., Verma A., Red Blood Cell Antibodies in Thalassemia Patients in Northern India: Risk Factors and Literature Review, *Indian Journal of Hematology and Blood Transfusion*, 2014, **30**:301 [Crossref], [Google Scholar], [Publisher]

[7]. Cappellini M.D., Cohen A., Eleftheriou A., Piga A., Porter J., Taher A., in *Guidelines for the Clinical Management of Thalassaemia [Internet]. 2nd Revised edition*, 2008 [Google Scholar], [Publisher]

[8]. Galanello R., Origa R., Beta-thalassemia, *Orphanet Journal of Rare Diseases*, 2010, **5**:11 [Crossref], [Google Scholar], [Publisher]

[9]. Tambunan B.A., Ugrasena I.D.G., Aryati., Role of Hemin in the Immune Response of T Follicular Helper Lymphocytes Expressing T-Cell Immunoreceptor with Immunoglobulin and Immunoreceptor **Tyrosine-Based** Inhibitory Domains, Programmed Cell Death-1, and Interleukin-21 in Allo-Auto Positive and Negative Thalassemia, Journal of Blood Medicine, 2023, **14**:7 [Google Scholar], [Publisher]

[10]. Sutrisnaningsih E.S., Suharjono S.,
Sudarmanto B., Analysis of Deferasirox and
Deferipron Use in Children with Pediatric βThalassemia Major, *Folia Medica Indonesiana*,
2017, **52**:42 [Crossref], [Google Scholar],
[Publisher]

[11]. Maharani E.A., Noviar G., Imunohematologi dan Bank Darah, *Pusat Pendidikan Sumber Daya Manusia Kesehatan, Badan Pengembangan dan Pemberdayaan Sumber Daya Manusia Kesehatan. Jakarta: Kementerian Kesehatan RI*, 2018 [Google Scholar], [Publisher]

[12]. Kurniawan A., Atmakusuma T.D., SukrismanL., Harimurti K., Failure on Maintaining

Haemoglobin Level After Transfusion in Transfusion Dependent Thalassemia Patiants Related to Erythrocyte Alloantibody and Autoantibody Production: Proportion and REL, 2013, [Crossref], [Google Scholar], [Publisher]

[13]. Essa E.S., El-Hawy M.A., Ahmedy I.A., Elmaghraby A.S., Red-blood cell alloimmunization in a cohort of multitransfused β-thalassemic patients in Menoufia Governorate, Egypt, *Menoufia Medical Journal*, 2022, **35**:34 [Crossref], [Google Scholar], [Publisher]

[14]. Singer S.T., Wu V., Mignacca R., Kuypers F.A., Morel P., Vichinsky E.P., Alloimmunization and erythrocyte autoimmunization in transfusiondependent thalassemia patients of predominantly Asian descent, *Blood, The Journal of the American Society of Hematology*, 2000, **96**:3369 [Crossref], [Google Scholar], [Publisher]

[15]. Ameen R., Al-Shemmari S., Al-Humood S., Chowdhury R.I., Al-Eyaadi O., Al-Bashir A., RBC alloimmunization and autoimmunization among transfusion-dependent Arab thalassemia patients, *Transfusion*, 2003, **43**:1604 [Crossref], [Google Scholar], [Publisher]

[16]. Dhawan H.K., Kumawat V., Marwaha N., Sharma R.R., Sachdev S., Bansal D., Marwaha R.K., Arora S., Alloimmunization and autoimmunization in transfusion dependent thalassemia major patients: Study on 319 patients, *Asian journal of transfusion science*, 2014, **8**:84 [Crossref], [Google Scholar], [Publisher]

[17]. Hendrickson J.E., Desmarets M., Deshpande S.S., Chadwick T.E., Hillyer C.D., Roback J.D., et al., Recipient inflammation affects the frequency and magnitude of immunization to transfused red blood cells, *Transfusion*, 2006, **46**:1526 [Crossref], [Google Scholar], [Publisher]

[18]. Saifeldeen E.R., Awad M.A., El-Tonbary Y.A., Aladle D.A., Elghannam D.M., Risk for red cell immunization among thalassemic patients, *The Egyptian Journal of Haematology*, 2017, **42**:58 [Crossref], [Google Scholar], [Publisher]

[19]. Shamsian B.S., Arzanian M.T., Shamshiri A.R., Alavi S., Khojasteh O., Frequency of Red Cell Alloimmunization in Patients with β-Major Thalassemia in an Iranian Referral Hospital, *Iranian Journal of Pediatric*, 2008, **18**:149 [Google Scholar], [Publisher] [20]. Ahmed A.M., Hasan N.S., Hassan Ragab S., Habib S.A., Emara N.A., Ahmed Aly A., Red cell alloimmunization and autoantibodies in Egyptian transfusion-dependent thalassaemia patients, *Archives of Medical Science*, 2010, **4**:592 [Crossref], [Google Scholar], [Publisher]

[21]. Obeidi N., Mankhian A.R., Hatami G., Emami
H., Antibody Screening in Patients With
Thalassemia Major, *Laboratory Medicine*, 2011,
42:618 [Crossref], [Google Scholar], [Publisher]

[22]. Vichinsky E., Neumayr L., Trimble S., Giardina P.J., Cohen A.R., Coates T., et al., Transfusion complications in thalassemia patients: a report from the <scp>C</scp> enters for <scp>D</scp> isease <scp>C</scp> ontrol and <scp>P</scp> revention (CME), *Transfusion*, 2014, **54**:972 [Crossref], [Google Scholar], [Publisher]

[23]. Pahuja S., Pujani M., Gupta S.K., Chandra J.,
Jain M., Alloimmunization and red cell autoimmunization in multitransfused thalassemics of Indian origin, *Hematology*, 2010,
15:174 [Crossref], [Google Scholar], [Publisher]

[24]. Wang L.Y., Liang D.C., Liu H.C., Chang F.C., Wang C.L., Chan Y.S., Lin M., Alloimmunization among patients with transfusion-dependent thalassemia in Taiwan, *Transfusion Medicine*, 2006, **16**:200 [Crossref], [Google Scholar], [Publisher]

[25]. Ghio M., Contini P., Mazzei C., Brenci S., Barberis G., Filaci G., Indiveri F., Puppo F., Soluble HLA class I, HLA class II, and Fas ligand in blood components: a possible key to explain the immunomodulatory effects of allogeneic blood transfusions, *Blood, The Journal of the American Society of Hematology*, 1999, **93**:1770 [Crossref], [Google Scholar], [Publisher]

[26]. Noor Haslina M.N., Ariffin N., Illuni Hayati I., Rosline H., Red cell autoantibodies among thalassaemia patients in Hospital Universiti Sains Malaysia., *Singapore medical journal*, 2007, 48:922 [Google Scholar], [Publisher]

[27]. Martelli A., Tazzari P., Bortul R., Riccio M., Tabellini G., Santi S., Frabetti F., Musiani D., Bareggi R., Conte R., Nuclear matrix protein is released from apoptotic white cells during cold (1-6° C) storage of concentrated red cell units and might induce antibody response in multiply transfused patients, *Transfusion*, 2000, **40**:169 [Crossref], [Google Scholar], [Publisher]

[28]. Pistillo M.P., Tazzari P.L., Gaudiano C., Cilla V., Kato T., Matsui T., Nishioka K., Capanni P., Conte R., Ferrara G.B., Patients with neoplastic and nonneoplastic hematologic diseases acquire CTLA-4 antibodies after blood transfusion, *Transfusion*, 2001, **41**:462 [Crossref], [Google Scholar], [Publisher]

[29]. Frabetti F., Musiani D., Marini M., Fanelli C., Coppola S., Ghibelli L., Tazzari P., Bontadini A., Tassi C., Conte R., White cell apoptosis in packed red cells, *Transfusion*, 1998, **38**:1082 [Crossref], [Google Scholar], [Publisher]

[30]. Hall A.M., Ward F.J., Vickers M.A., Stott L.M., Urbaniak S.J., Barker R.N., Interleukin-10– mediated regulatory T-cell responses to epitopes on a human red blood cell autoantigen, *Blood*, 2002, **100**:4529 [Crossref], [Google Scholar], [Publisher]

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